

Research Article

Variation in chemical constituents of essential oils of the fresh, dried and fermented leaves of *Premna serratifolia*

Khin Su Yee^{1,2}, Penpun Wetwitayaklung¹, Worrakanya Narakornwit¹,
Tasamaporn Sukwattanasinit¹, Bunyapa Wangwattana¹, Uthai Sotanaphun^{1*}

¹ Department of Pharmacognosy, Faculty of Pharmacy, Silpakorn University, Nakhon Pathom, Thailand

² Department of Pharmacognosy, University of Pharmacy, Mandalay, Myanmar

ABSTRACT

This study aimed to identify the composition of the essential oils of the fresh, dried and fermented leaves of *Premna serratifolia* by gas chromatography-mass spectrometry (GC-MS) using DB-5 and Carbowax 20M columns. A total of 77, 82 and 90 compounds were detected, which involved the main compound categories of hydrocarbons, terpenoids, and phenolics. Amyl vinyl carbinol (15.8-32.6%), linalool (11.1-15.1%), phytol (7.7-12.5%), salicylic acid methyl ester (3.9-7.2%) and (*E*)-caryophyllene (3.1-6.6%) were predominant components of the fresh leaf oils. After drying and fermentation, the chemical compositions were changed by various reactions. The amounts of amyl vinyl carbinol were decreased to 6.3-13.8% and 6.9-11.5% after drying and fermentation, respectively. Likewise, linalool and phytol were decreased to 6.3-7.5% and 7.3-9.0% after drying, and decreased to 5.3-7.9% and 2.0-3.4% after fermentation, respectively. In both the dried and the fermented leaf oils, alpha-humulene was disappeared and beta-myrcene was detected as a new compound. The noticeable changes in chemical composition after drying process were much increasing in the amount of (*E*)-caryophyllene (6.6-12.2%) to become the most abundant compound, and the hydrolysis of palmitic acid ethyl ester to palmitic acid (5.0%). The fermentation method could dramatically increase the amounts of phenolic compounds especially *p*-vinylanisole (2.4-41.1%) which became the major compound, marked decrease the phytol and found acorenone B (4.4%) as a new compound. The present study demonstrated that drying and fermentation processes affected the volatile composition of the leaves of *P. serratifolia*.

Keywords:

Premna serratifolia, Essential oil, GC-MS, Drying, Fermentation

1. INTRODUCTION

Premna serratifolia L. (family Lamiaceae) is a shrub or tree in tropical and subtropical regions. Its leaves have a characteristic fetid smell and native people in Celebes use it as a food additive to reduce the fishy smell¹. In Peninsula, Malaysia, and Indonesia, young leaves of this plant are used as vegetables². In Myanmar, the leaves were used for the treatments of cancer and liver diseases. Some people take the preparation of the fresh leaves and some others use the dried leaves. If the fresh leaves of this plant were kept at room temperature or dried in the shade under inappropriate condition, some leaves will be fermented and turned to dark brown

color. The fresh, dried and fermented leaves have different smells which suggested that their volatile chemical constituents should be in variation. Many medicinal plants have been reported for chemical variation between their fresh and dried leaves, such as *Tapinanthus bangwensis*³, *Artemisia afra*⁴, *Ocimum sanctum*⁵, and *Cymbopogon citratus*⁶. One of the most well-known fermented example was the hydrolysis of aroma precursors by endogenous glycosidase during the manufacturing processes of Oolong tea and black tea, and many aroma compounds occurred⁷.

This study was a keen interest to investigate the difference among volatile phytochemical constituents of the fresh, dried and fermented leaves of *P. serratifolia*

*Corresponding author:

*Uthai Sotanaphun sotaphun_u@su.ac.th



Pharmaceutical Sciences Asia © 2021 by

Faculty of Pharmacy, Mahidol University, Thailand is licensed under CC BY-NC-ND 4.0. To view a copy of this license, visit <https://www.creativecommons.org/licenses/by-nc-nd/4.0/>

using gas chromatography coupled with mass spectrometry (GC-MS) technique. The result would be declared as basic scientific evidence that their qualities were not equal and should be concerned.

2. MATERIALS AND METHODS

2.1. Plant material

Leaves of *P. serratifolia* were collected from Mandalay Division, Myanmar, in October 2018. The taxonomic species was identified by comparing its flower and leaf characters with the references⁸⁻¹⁰. The voucher specimen (Ps172018) was deposited with the herbarium of the Department of Pharmacognosy, Silpakorn University, Thailand. The leaves were divided into three parts. The first part was the fresh leaves, the second part was dried in the shade, and the third part was fermented by packing in a plastic bag and place at room temperature for 10 days in the shade until the leaves turned to black color.

2.2. Preparation of essential oils

The fresh leaf sample (490 g), the dried leaf sample (244 g) and the fermented leaf sample (201 g) were separately chopped into small pieces and hydro-distilled in a Clevenger apparatus for 5 hours to obtain essential oils. The collected oils were dried over anhydrous sodium sulfate and stored at 4°C in air-tight glass containers. The yields of the essential oils of the fresh, dried and fermented leaves were 0.008, 0.139 and 0.139%, respectively.

2.3. GC-MS analysis

The essential oils were analyzed by Agilent 6890 gas chromatography equipped with Agilent technology, 5973N mass selective spectrometric detector (EIMS, electron energy, 70 eV, scanning from 40 to 500 m/z) with a quadrupole analyzer and an Agilent Chem Station data system (Agilent Technologies, U.S.A.). Two columns, fused silica capillary column (5%-phenyl)-methylpolysiloxane DB-5 (30 m x 0.32 mm ID x 0.25 µm film thickness) and Carbowax 20M polyethylene glycol (PEG) (60 m x 0.25 mm ID x 0.25 µm film thickness) were used. Ultra-high purity helium gas (99.999%) was used as a carrier gas at a flow rate of 1 mL/min. The sample (1.0 µL) was injected with a splitless mode. The solvent delay for the detector was 3 minutes. The ion source temperature was 230°C and quadrupole temperature was programmed at 150°C. For the DB-5 column, the initial oven temperature was 80°C and increased to 130°C at the 5°C/min, then increased to 280°C at the 10°C/min and hold for 5 min. For Carbowax 20M column, the initial oven tempe-

rate was 60°C and increased to 220°C at the 2°C/min. Identification of compounds was performed by comparison of their RI relative to *n*-alkanes (C8-C26) with Adams¹¹, NIST Chemistry WebBook, SRD 69¹², Babushok *et al*¹³ and Leffingwell *et al*¹⁴. Their mass spectra were also compared with libraries databases of Wiley7n.l and NIST 05.

3. RESULTS AND DISCUSSION

Essential oils of the fresh leaves of *P. serratifolia* were prepared by hydrodistillation and studied by GC-MS. Two GC columns, the non-polar DB-5, and the polar Carbowax 20M columns were used to ensure the most complete investigation of the constituents. The identification of each compound was based on its mass fragmentation pattern and RI (Retention Index) calculation comparing with data in the references. The results are shown in Table 1 and Table 2. Seventy-seven compounds were detected. Most of the identified compounds were classified into hydrocarbons (26.1% in DB-5 and 57.1% in Carbowax 20M), terpenoids (42.8% in DB-5 and 26.0% in Carbowax 20M) and phenolics (15.0% in DB-5 and 10.9% in Carbowax 20M) compound categories. Some fatty acids, apocarotenoids, and miscellaneous compounds were also detected (Table 3). The most abundant compounds were amyl vinyl carbinol (15.8% in DB-5 and 32.6% in Carbowax 20M), linalool (15.1% in DB-5 and 11.1% in Carbowax 20M), phytol (12.5% in DB-5, and 7.7% in Carbowax 20M), salicylic acid methyl ester (7.2% in DB-5 and 3.9% in Carbowax 20M) and (*E*)-caryophyllene (6.6% in DB-5 and 3.1% in Carbowax 20M). The previous study identified only 4 compounds of eugenol, eugenyl acetate, massoil and *cis*-2-oxabicyclo, 4.4.0-decane in the fresh leaf oil¹. In this study, a very less amount of eugenol was detected (1.2% in DB5). This might be due to the different location and climate of plant origin. This study was the first report of the constituent of essential oil obtained from the leaves of this plant growing in Myanmar.

After the leaves of *P. serratifolia* were dried under the shade, increasing in the number of compounds in essential oils was observed, resulting in 82 total compounds; whereas when the leaves were allowed to ferment and turn to dark color, 90 compounds were detected (Tables 1 and Table 2). Some compounds were disappeared and some new compounds occurred as concluded in Figure 1. The compounds those lose after drying and fermentation methods were not much different. The significantly lost compounds were alpha-humulene, amyl carbinol, (*Z*)-3-hexen-1-ol and acetophenone. The new occurring compounds were mostly the small molecular weight terpenoids and aldehyde hydrocarbons compound categories, but they were detected in only trace amounts.

Table 1. Chemical constituents of the essential oils of the fresh, dried and fermented *P. serratifolia* leaves analysed by GC-MS using DB-5 column.

| Compound | Fresh leaves | | Dried leaves | | Fermented leaves | | Literature RI |
|--|--------------|---------------------|--------------|---------------------|------------------|---------------------|--------------------|
| | RI | %Relative amount | RI | %Relative amount | RI | %Relative amount | |
| alpha-Pinene | - | - | 1015 | 0.8 | - | - | 939 ¹¹ |
| Amyl vinyl carbinol | 1031 | 15.8 | 1030 | 6.3 | 1031 | 6.9 | 979 ¹¹ |
| Amyl ethyl ketone | - | - | - | - | 1034 | 1.2 | 984 ¹¹ |
| beta-Myrcene | - | - | 1037 | 8.7 | 1037 | 6.7 | 991 ¹¹ |
| Amyl ethyl carbinol | 1049 | 6.4 | - | - | 1040 | 5.7 | 1002 ¹² |
| Unknown | - | - | - | - | 1048 | 0.7 | - |
| <i>m</i> -cymene | 1059 | 0.2 | - | - | - | - | 1026 ¹² |
| Limonene | 1069 | 0.3 | 1065 | 0.3 | 1066 | 0.4 | 1029 ¹¹ |
| Benzeneacetaldehyde | 1079 | 1.5 | - | - | 1076 | 1.0 | 1042 ¹¹ |
| Unknown | - | - | - | - | 1115 | 1.2 | - |
| Linalool | 1126 | 15.1 | 1122 | 6.3 | 1123 | 7.9 | 1097 ¹¹ |
| (<i>Z</i>)-beta-Terpineol | - | - | - | - | 1125 | 0.6 | 1144 ¹¹ |
| Unknown | - | - | - | - | 1144 | 0.4 | - |
| <i>p</i> -Vinyl Anisole | 1172 | 4.5 | 1172 | 8.9 | 1184 | 1.4 | 1160 ¹² |
| <i>neo</i> -Menthol | - | - | 1190 | 0.6 | - | - | 1166 ¹¹ |
| Unknown | - | - | - | - | 1190 | 1.1 | - |
| Naphthalene | 1204 | 0.6 | - | - | - | - | 1181 ¹¹ |
| Unknown | - | - | - | - | 1205 | 0.5 | - |
| alpha-Terpineol | - | - | - | - | 1208 | 0.7 | 1189 ¹¹ |
| Unknown | - | - | 1208 | 5.9 | - | - | - |
| Salicylic acid methyl ester | 1216 | 7.2 | - | - | - | - | 1193 ¹³ |
| beta-Cyclocitral | 1236 | 0.7 | - | - | - | - | 1218 ¹³ |
| Nerol | 1239 | 0.5 | 1237 | 0.3 | - | - | 1230 ¹¹ |
| 5-(1'-1'-Dimethylethyl)bicycle[3,10] hexan-2-one | - | - | - | - | 1256 | 1.7 | - |
| <i>p</i> -Anisaldehyde | - | - | - | - | 1260 | 1.9 | 1270 ¹² |
| Unknown | - | - | 1270 | 1.0 | - | - | - |
| alpha-Benzeneacetaldehyde ethylidene- | - | - | 1282 | 0.2 | 1279 | 0.8 | 1279 ¹² |
| Salicylic acid ethyl ester | 1282 | 1.1 | - | - | - | - | 1311 ¹² |
| Carbamic acid | - | - | 1307 | 0.7 | - | - | - |
| 3,4,4a,5,6,8a-Hexahydro-2,5,5,8a-tetramethyl- (2.alpha.,4a.alpha.,8a.alpha.)-2H-1-benzopyran | 1307 | 0.5 | - | - | - | - | - |
| <i>p</i> -Vinylgylacol | 1323 | 1.1 | 1322 | 0.1 | 1322 | 0.6 | 1324 ¹² |
| Benzene,4-ethyl-1,2-dimethoxy- | - | - | - | - | 1329 | 0.6 | - |
| Unknown | - | - | - | - | 1332 | 0.7 | - |
| Acetanisole | - | - | 1361 | 0.1 | 1362 | 0.8 | 1352 ¹¹ |
| Eugenol | 1366 | 1.2 | 1365 | 0.7 | 1366 | 1.7 | 1359 ¹¹ |
| Unknown | 1369 | 1.3 | 1368 | 0.6 | - | - | - |
| 3,4-Dimethoxystyrene | - | - | - | - | 1371 | 0.6 | 1368 ¹² |
| Unknown | - | - | - | - | 1384 | 1.4 | - |
| (<i>E</i>)-beta-Damascenone | 1392 | 1.4 | 1392 | 0.7 | - | - | 1385 ¹¹ |
| beta-Elementene | 1400 | 0.3 | 1400 | 0.4 | 1400 | 0.6 | 1391 ¹¹ |
| Methyleugenol | - | - | 1407 | 0.4 | 1407 | 0.9 | 1404 ¹¹ |
| Unknown | - | - | 1413 | 0.4 | - | - | - |
| Isocaryophyllene | - | - | - | - | 1420 | 1.1 | 1423 ¹² |
| Unknown | - | - | 1421 | 0.7 | - | - | - |
| (<i>E</i>)-Caryophyllene | 1436 | 6.6 | 1438 | 12.2 | 1437 | 12.7 | 1433 ¹² |
| Dihydro-beta-ionone | - | - | 1447 | 0.4 | - | - | 1444 ¹² |
| Neryl acetone | - | - | 1456 | 1.0 | 1456 | 0.6 | 1456 ¹² |
| (<i>E</i>)-beta-Farnesene | 1459 | 0.4 | 1460 | 0.5 | 1460 | 0.6 | 1457 ¹¹ |
| alpha-Humulene | 1467 | 1.6 | - | - | - | - | 1463 ¹² |
| 1,1,4,8-Tetramethyl-(<i>Z,Z,Z</i>)-4,7,10-cycloundecatriene- | - | - | 1468 | 2.4 | 1468 | 3.0 | - |
| (<i>E</i>)-beta-Ionone | 1491 | 0.9 | 1493 | 2.9 | 1493 | 1.7 | 1489 ¹¹ |
| Unknown | - | - | - | - | 1499 | 1.1 | - |
| alpha-Farnesene | 1509 | 0.2 | - | - | - | - | 1506 ¹² |
| beta-Bisabolene | 1514 | 3.0 | 1515 | 3.8 | 1515 | 4.8 | 1512 ¹² |
| Unknown | - | - | 1529 | 0.8 | 1526 | 1.4 | - |
| beta-Cadinene | - | - | - | - | 1533 | 1.1 | 1539 ¹¹ |
| Unknown | - | - | 1534 | 1.4 | - | - | - |
| Nerolidol | 1566 | 0.9 | 1566 | 1.3 | 1567 | 1.3 | 1563 ¹¹ |

Table 1. Chemical constituents of the essential oils of the fresh, dried and fermented *P. serratifolia* leaves analysed by GC-MS using DB-5 column. (cont.)

| Compound | Fresh leaves | | Dried leaves | | Fermented leaves | | Literature RI |
|--|--------------|---------------------|--------------|---------------------|------------------|---------------------|--------------------|
| | RI | %Relative amount | RI | %Relative amount | RI | %Relative amount | |
| Caryophyllene oxide | 1597 | 1.8 | 1598 | 2.6 | 1598 | 2.9 | 1583 ¹¹ |
| Humulene oxide | - | - | - | - | 1628 | 0.4 | 1602 ¹³ |
| 3,4-Dimethyl-3-cyclohexen-1-carboxaldehyde | 1629 | 0.3 | - | - | - | - | - |
| beta-Turmerone | - | - | 1674 | 1.0 | - | - | 1669 ¹¹ |
| Unknown | - | - | - | - | 1678 | 2.4 | - |
| Unknown | - | - | - | - | 1681 | 2.7 | - |
| Acorenone B | - | - | - | - | 1708 | 4.4 | 1698 ¹¹ |
| Myristic acid ethyl ester | 1794 | 0.3 | - | - | - | - | 1794 ¹² |
| Isopropyl myristate | 1826 | 0.3 | - | - | - | - | 1824 ¹² |
| Hexahydrofarnesyl acetone | 1848 | 0.3 | 1844 | 3.6 | 1849 | 1.5 | 1844 ¹² |
| Isobutyl phthalate | 1875 | 0.2 | - | - | 1875 | 0.3 | 1874 ¹² |
| Palmitoleic acid methyl ester | - | - | 1912 | 0.9 | 1911 | 0.3 | 1912 ¹² |
| Farnesyl acetone C | 1925 | 0.4 | 1926 | 2.0 | 1925 | 0.7 | 1920 ¹³ |
| Isophytol | - | - | - | - | 1951 | 0.1 | 1948 ¹¹ |
| (Z)-11-Hexadecenoic acid | - | - | 1958 | 1.8 | - | - | 1953 ¹² |
| Unknown | 1964 | 1.3 | 1963 | 1.0 | 1963 | 0.8 | - |
| Palmitic acid | - | - | 1978 | 5.0 | - | - | 1968 ¹³ |
| (E)-11-Hexadecenoic acid ethyl ester | 1980 | 0.8 | - | - | 1979 | 0.4 | 1974 ¹² |
| Palmitic acid ethyl ester | 1990 | 1.2 | - | - | - | - | 1990 ¹² |
| Geranyl linalool isomer1 | - | - | 2038 | 0.2 | - | - | - |
| Linoleic acid methyl ester | - | - | 2098 | 0.4 | - | - | 2097 ¹² |
| Linolenic acid methyl ester | - | - | 2106 | 1.2 | 2105 | 0.3 | 2108 ¹² |
| Phytol | 2128 | 12.5 | 2124 | 7.3 | 2120 | 3.4 | 2122 ¹² |
| (E)-9-Octadecenoic acid | - | - | 2146 | 0.4 | - | - | 2133 ¹³ |
| Linoleic acid ethyl ester | 2167 | 0.6 | - | - | 2165 | 0.1 | 2164 ¹² |
| Linolenic acid ethyl ester | 2175 | 2.3 | 2174 | 0.4 | 2173 | 0.6 | 2170 ¹² |
| Geranyl linalool isomer2 | - | - | 2180 | 0.4 | - | - | - |
| 15-Methyl-heptadecanoic acid ethyl ester | 2194 | 0.4 | - | - | - | - | - |
| n-Docosane | - | - | - | - | 2199 | 0.1 | 2200 ¹⁴ |
| n-Tricosane | 2299 | 0.2 | 2299 | 0.1 | 2299 | 0.1 | 2300 ¹⁴ |
| n-Tetracosane | 2399 | 0.1 | 2399 | 0.1 | 2399 | 0.1 | 2400 ¹⁴ |
| n-Pentacosane | 2499 | 0.2 | 2499 | 0.1 | 2499 | 0.2 | 2500 ¹⁴ |
| Bis-(2-ethylhexyl) phthalate | 2554 | 3.0 | 2552 | 0.1 | - | - | 2556 ¹⁴ |
| n-Hexacosane | 2599 | 0.1 | 2598 | 0.1 | 2598 | 0.2 | 2600 ¹⁴ |
| n-Heptacosane | 2700 | 0.4 | 2699 | 0.4 | 2698 | 0.3 | 2700 ¹⁴ |
| Squalene | 2826 | 0.2 | - | - | 2825 | 0.1 | 2847 ¹² |
| n-Nonacosane | 2897 | 3.0 | 2898 | 0.4 | 2898 | 0.4 | 2900 ¹⁴ |

Table 2. Chemical constituents of the essential oils of the fresh, dried and fermented *P. serratifolia* leaves analysed by GC-MS using Carbowax 20M column.

| Compound | Fresh leaves | | Dried leaves | | Fermented leaves | | Literature RI |
|--|--------------|---------------------|--------------|---------------------|------------------|---------------------|--------------------|
| | RI | %Relative amount | RI | %Relative amount | RI | %Relative amount | |
| (E)-2-Hexenal | 1271 | 0.6 | 1278 | 0.7 | 1279 | 0.3 | 1216 ¹² |
| (E)-beta-Ocimene | - | - | 1282 | 0.2 | 1283 | 0.2 | 1250 ¹² |
| Amyl ethyl ketone | 1299 | 7.8 | 1297 | 0.3 | 1298 | 0.7 | 1264 ¹² |
| p-Cymene | - | - | 1307 | 0.2 | 1300 | 0.2 | 1270 ¹³ |
| alpha-Terpinolen | - | - | 1308 | 0.1 | - | - | 1282 ¹³ |
| Amyl vinyl ketone | 1325 | 0.3 | 1326 | 1.4 | 1326 | 0.2 | 1301 ¹³ |
| 6-Methyl-5-hepten-2-one | - | - | 1355 | 0.1 | 1355 | 0.1 | 1345 ¹² |
| Amyl carbinol | 1379 | 5.4 | - | - | - | - | 1371 ¹² |
| (Z)-3-Hexen-1-ol | 1401 | 2.2 | - | - | - | - | 1373 ¹³ |
| Amyl ethyl carbinol | 1423 | 8.1 | 1414 | 2.1 | 1416 | 6.2 | 1392 ¹³ |
| 3,5,5-Trimethyl-3-cyclohexen-1-one | - | - | 1425 | 0.1 | - | - | 1420 ¹² |
| Amyl vinyl carbinol | 1474 | 32.6 | 1460 | 13.8 | 1461 | 11.5 | 1444 ¹³ |
| (E)-Linalool oxide | 1476 | 0.2 | - | - | - | - | 1454 ¹³ |
| (Z)-Linalool oxide | 1491 | 0.2 | 1486 | 0.2 | 1486 | 0.3 | 1474 ¹³ |
| (E,E)-2,4-Heptadienal | 1502 | 0.1 | 1498 | 0.2 | 1499 | 0.2 | 1491 ¹³ |
| 3,4,4a,5,6,8a-Hexahydro-2,5,5,8a-tetramethy-(2.alpha.,4a.alpha.,8a.alpha.) 2H-1-benzopyran | 1527 | 0.2 | - | - | - | - | - |

Table 2. Chemical constituents of the essential oils of the fresh, dried and fermented *P. serratifolia* leaves analysed by GC-MS using Carbowax 20M column. (cont.)

| Compound | Fresh leaves | | Dried leaves | | Fermented leaves | | Literature RI |
|--|--------------|---------------------|--------------|---------------------|------------------|---------------------|--------------------|
| | RI | %Relative amount | RI | %Relative amount | RI | %Relative amount | |
| Camphor | - | - | 1531 | 0.1 | - | - | 1515 ¹³ |
| (<i>E</i>)-2-Nonenal | - | - | 1538 | 0.2 | 1540 | 0.2 | 1536 ¹³ |
| Linalool | 1554 | 11.1 | 1549 | 7.5 | 1549 | 5.3 | 1543 ¹³ |
| 1-Methyl-4-(1-methylethyl)- <i>trans</i> -2-cyclohexen-1-ol | - | - | - | - | 1577 | 0.3 | 1571 ¹² |
| beta-Elementene | - | - | 1578 | 0.2 | - | - | 1574 ¹² |
| (<i>E,Z</i>)-2,6-Nonadienal | - | - | 1583 | 0.4 | 1585 | 0.6 | 1582 ¹³ |
| (<i>E</i>)-Caryophyllene | 1594 | 3.1 | 1596 | 14.6 | 1597 | 8.4 | 1599 ¹³ |
| Unknown | - | - | - | - | 1603 | 0.6 | - |
| 2-Acetylthiazole | 1632 | 0.1 | - | - | - | - | 1634 ¹² |
| Acetophenone | 1645 | 2.0 | - | - | - | - | 1648 ¹² |
| (<i>E</i>)-beta-Farnesene | - | - | - | - | 1651 | 0.3 | 1664 ¹³ |
| <i>p</i> -Vinyl anisole | 1659 | 2.4 | 1660 | 13.1 | 1667 | 41.1 | 1670 ¹² |
| Unknown | 1667 | 0.2 | - | - | 1671 | 0.6 | - |
| 4-Oxoisophorone | - | - | 1694 | 0.2 | 1696 | 0.2 | 1690 ¹² |
| (<i>E,E</i>)-2,4 Nonadienal | - | - | - | - | 1698 | 0.1 | 1696 ¹³ |
| beta-Bisabolene | 1706 | 1.5 | 1707 | 6.0 | 1706 | 2.7 | 1699 ¹² |
| Naphthalene | 1717 | 0.2 | 1717 | 0.2 | - | - | 1709 ¹² |
| alpha-Farnesene | - | - | 1722 | 0.4 | 1722 | 0.2 | 1744 ¹³ |
| Unknown | 1722 | 0.2 | - | - | - | - | - |
| delta-Cadinene | - | - | 1737 | 0.2 | - | - | 1756 ¹³ |
| Salicylic acid methyl ester | 1755 | 3.9 | 1751 | 0.5 | 1752 | 0.3 | 1768 ¹³ |
| Salicylic acid ethyl ester | 1788 | 0.4 | - | - | - | - | 1798 ¹² |
| Nerol | - | - | 1802 | 0.3 | 1801 | 0.1 | 1795 ¹³ |
| Unknown | 1802 | 0.4 | - | - | - | - | - |
| (<i>E</i>)-beta-Damascenone | 1814 | 0.6 | 1813 | 0.7 | 1813 | 0.2 | 1821 ¹³ |
| Dihydro-beta-Ionone | - | - | 1827 | 0.3 | - | - | 1825 ¹² |
| Unknown | 1830 | 0.2 | - | - | - | - | - |
| Neryl acetone | 1843 | 0.1 | 1844 | 1.2 | 1844 | 0.4 | 1835 ¹² |
| Geraniol | 1852 | 0.7 | 1851 | 0.7 | 1851 | 0.4 | 1851 ¹² |
| alpha-Ionone | - | - | 1853 | 0.6 | - | - | 1857 ¹² |
| 4-Ethyl-1,2-dimethoxy-benzene | - | - | - | - | 1870 | 0.4 | 1875 ¹² |
| Unknown | 1877 | 0.6 | 1876 | 0.7 | 1877 | 1.1 | - |
| (<i>E,E,E</i>)-2,4,6-Nona-trienal | - | - | 1889 | 0.2 | 1889 | 0.4 | - |
| Unknown | 1913 | 0.2 | - | - | - | - | - |
| Unknown | 1923 | 0.2 | - | - | 1923 | 0.5 | - |
| (<i>E</i>)-beta-Ionone | 1943 | 0.2 | 1945 | 2.2 | 1944 | 0.4 | 1936 ¹³ |
| Unknown | 1978 | 0.2 | - | - | - | - | - |
| Caryophyllene oxide | 1991 | 0.7 | 1992 | 2.5 | 1991 | 1.0 | 1986 ¹³ |
| Methyleugenol | - | - | 2002 | 0.7 | 2002 | 0.5 | 2006 ¹³ |
| 4-(2,2,6-Trimethyl-7-oxabicyclo[4.1.0]hept-1-yl)-3-buten-2-one | 2002 | 0.1 | - | - | - | - | 2002 ¹² |
| <i>p</i> -Anisaldehyde | 2020 | 0.5 | 2020 | 1.7 | 2021 | 1.5 | 2011 ¹³ |
| Nerolidol | 2029 | 0.4 | 2029 | 1.4 | 2029 | 0.4 | 2036 ¹³ |
| Myristic acid ethyl ester | 2037 | 0.1 | - | - | - | - | 2045 ¹² |
| Zingiberenol | - | - | - | - | 2114 | 0.1 | 2109 ¹² |
| Hexahydro farnesyl acetone | 2121 | 0.1 | 2124 | 4.1 | 2122 | 0.8 | 2129 ¹² |
| Unknown | - | - | - | - | 2100 | 0.2 | - |
| Acetanisole | 2144 | 0.1 | 2144 | 0.4 | 2145 | 0.7 | 2148 ¹² |
| Eugenol | 2151 | 0.6 | 2151 | 1.5 | 2151 | 1.1 | 2150 ¹² |
| Longiborneol | - | - | 2160 | 0.4 | 2159 | 0.2 | 2157 ¹³ |
| 2,6,11,15-Tetramethyl-hexadeca-2,6,8,10,14-pentaene | 2175 | 0.6 | 2174 | 1.1 | 2174 | 0.4 | - |
| <i>p</i> -Vinylguaiaicol | 2184 | 0.6 | - | - | 2184 | 0.4 | 2180 ¹² |
| alpha-Cadinol | 2191 | 0.1 | 2191 | 0.3 | - | - | 2191 ¹² |
| Unknown | - | - | 2202 | 1.7 | 2202 | 0.5 | - |
| Acorenone B | - | - | - | - | 2217 | 2.4 | - |
| Palmitoleic acid methyl ester | 2226 | 0.1 | 2227 | 0.9 | - | - | 2225 ¹² |
| Palmitic acid ethyl ester | 2245 | 0.4 | - | - | 2243 | 0.2 | 2235 ¹² |
| beta-Turmerone | - | - | 2246 | 0.5 | - | - | - |
| 9-hexadecenoic ethyl ester | 2268 | 0.3 | - | - | 2266 | 0.2 | 2269 ¹² |
| Unknown | - | - | - | - | 2281 | 1.7 | - |

Table 2. Chemical constituents of the essential oils of the fresh, dried and fermented *P. serratifolia* leaves analysed by GC-MS using Carbowax 20M column. (cont.)

| Compound | Fresh leaves | | Dried leaves | | Fermented leaves | | Literature RI |
|---|--------------|------------------|--------------|------------------|------------------|------------------|--------------------|
| | RI | %Relative amount | RI | %Relative amount | RI | %Relative amount | |
| Isophytol | - | - | 2284 | 0.3 | - | - | 2293 ¹³ |
| 2,4-Bis(1,1-dimethylethyl)-phenol | 2291 | 0.1 | 2291 | 0.6 | 2291 | 0.2 | 2316 ¹² |
| 3,7,11-Trimethyl-(<i>E,E</i>)-2,6,10-dodecatrien-1-ol | - | - | - | - | 2357 | 0.1 | 2366 ¹³ |
| Farnesyl acetone C | 2362 | 0.1 | 2363 | 1.2 | 2362 | 0.4 | 2377 ¹³ |
| Ketole (1H-Indole) | - | - | 2425 | 0.2 | 2425 | 0.2 | 2420 ¹² |
| Octadecanoic acid ethyl ester | 2453 | 0.1 | - | - | - | - | 2450 ¹² |
| Oleic acid ethyl ester | 2468 | 0.2 | - | - | - | - | 2461 ¹² |
| Linoleic acid methyl ester | - | - | 2478 | 0.3 | - | - | 2480 ¹² |
| Linoleic acid ethyl ester | 2515 | 0.2 | - | - | - | - | 2519 ¹² |
| Linolenic acid methyl ester | - | - | 2547 | 1.1 | 2546 | 0.2 | 2550 ¹² |
| Linolenic acid ethyl ester | 2584 | 0.7 | - | - | 2582 | 0.4 | 2594 ¹² |
| Phytol | 2625 | 7.7 | 2620 | 9.0 | 2617 | 2.0 | 2613 ¹³ |
| Dibutyl phthalate | - | - | 2704 | 0.5 | - | - | 2705 ¹² |
| 3-(4-Methoxyphenyl),2-propenoic acid ethyl ester | 2641 | 0.5 | - | - | - | - | - |

Table 3. Category of chemical constituents of the essential oils of the fresh, dried and fermented *P. serratifolia* leaves.

| Compound category | Compound sub-category | % Relative amount | | | | | |
|-------------------|---|-------------------|--------------|--------------|--------------|------------------|--------------|
| | | Fresh leaves | | Dried leaves | | Fermented leaves | |
| | | DB-5 | Carbowax 20M | DB-5 | Carbowax 20M | DB-5 | Carbowax 20M |
| Terpenoids | Monoterpene hydrocarbons | 0.5 | - | 9.7 | 0.4 | 7.1 | 0.4 |
| | Monoterpene alcohols | 15.6 | 11.8 | 7.1 | 8.5 | 9.2 | 6.1 |
| | Monoterpene ketones | - | - | - | 0.1 | - | - |
| | Miscellaneous oxygenated monoterpenes | - | 0.4 | - | 0.2 | - | 0.3 |
| | Sesquiterpene hydrocarbons | 12.1 | 4.7 | 19.3 | 21.4 | 23.8 | 11.5 |
| | Sesquiterpenes alcohols | - | 0.1 | - | 0.7 | - | 0.3 |
| | Sesquiterpene ketones | - | - | 1.0 | 0.5 | 4.4 | 2.4 |
| | Miscellaneous oxygenated sesquiterpenes | 1.8 | 0.7 | 2.6 | 2.5 | 3.2 | 1.0 |
| | Diterpenoid hydrocarbons | - | 0.6 | - | 1.1 | - | 0.4 |
| | Diterpenoid alcohols | 12.5 | 7.7 | 7.9 | 9.3 | 3.5 | 2.0 |
| | Triterpenoids | 0.2 | - | - | - | 0.1 | - |
| Phenolics | | 15.0 | 10.9 | 10.2 | 18.4 | 7.9 | 6.0 |
| Fatty acids | Fatty acids | - | - | 7.2 | - | - | - |
| | Fatty acids esters | 5.7 | 2.1 | 2.9 | 2.2 | 1.7 | 1.0 |
| Hydrocarbons | Hydrocarbon alcohols | 22.2 | 48.2 | 6.3 | 15.9 | 12.5 | 17.8 |
| | Hydrocarbon aldehydes | - | 0.7 | - | 1.7 | - | 1.7 |
| | Hydrocarbon ketones | - | 8.1 | - | 1.6 | 2.9 | 0.9 |
| | Long chain hydrocarbons | 3.9 | - | 1.2 | - | 1.2 | - |
| Apocarotenoids | Apocarotenoids | 4.9 | 1.7 | 11.9 | 12.1 | 5.7 | 2.8 |
| Miscellaneous | | 2.8 | 0.2 | 0.9 | 0.9 | 2.7 | 0.2 |
| Unknowns | | 2.6 | 2.1 | 11.8 | 2.3 | 13.9 | 5.2 |

The significant new abundant compounds of both the dried and the fermented leaf oils were beta-myrcene and (*Z,Z,Z*)-4,7,10-cycloundecatriene,1,1,4,8-tetramethyl. The major difference between the dried and the fermented leaf oils was the detection of fatty acids (especially palmitic acid) only in the dried leaf oil, whereas acorenone B was found only in the fermented leaf oil.

Changing in a relative amount of each constituent was the major observation after drying or fermentation (Figure 2). Comparing with essential oil of the fresh leaves, essential oils of the dried leaves

and the fermented leaves possessed less amounts of monoterpenes (especially linalool), *vice versa*, a higher proportion of sesquiterpenes (e.g. (*E*)-caryophyllene, beta-bisabolene, caryophyllene oxide) were observed, especially (*E*)-caryophyllene that became the most abundant compound of the dried leaf oil (6.6-12.2% in DB-5 and 3.1-14.6% in Carbowax 20M). The amount of phytol, the only major diterpenoid compound of the fresh leaf oil, was dramatically decreased after fermentation, but changing of this compound after drying was not much significant. In overview, the amount of phenolic compounds did not change much after drying

while there was a marked increase after fermentation. In detail, the amount of salicylic acid methyl ester decreased in both the dried and the fermented leaf oils, but the amount of *p*-vinyl anisole was dramatically increased after fermentation more than the drying method and became the most abundant compound of the fermented leaf oil (2.4-41.1% Carbowax 20M). On the other hand, free fatty acids were found only in the dried leaf oil. In overview, the amounts of fatty acid esters (e.g. palmitic acid ethyl ester, linolenic acid ethyl ester) and hydrocarbons (e.g. amyl ethyl carbinol, amyl vinyl carbinol, amyl ethyl ketone) tended to decrease in both the dried and the fermented leaf oils.

The relation between chemical structures of some constituents of the essential oils of the leaves of *P. serratifolia* after drying or fermentation could be explained via different reactions. beta-Myrcene which was the new detected compound of both the dried and fermented leaf oils, possesses the same chemical skeleton of acyclic monoterpenoids as linalool that concomitant dramatically decreasing in relative amount. Dehydration of linalool to form beta-myrcene might be the mechanism of this reaction but it has never been reported in the plant. There was only a report found that linalool could be converted to beta-myrcene by microbial biotransformation using linalool dehydratase isomerase¹⁵. After drying and fermentation, alpha-humulene was disappeared, whereas relative amounts of (*E*)-caryophyllene and caryophyllene oxide increased. alpha-Humulene and (*E*)-caryophyllene possess a similar sesquiterpenoid skeleton and have humulyl cation as the same biosynthetic precursor¹⁶. Some unproven factors might affect their biosynthesis expression, and (*E*)-caryophyllene could be further oxidized to caryophyllene oxide¹⁷. Most of the fatty acid esters were found mainly in the fresh leaf oil (e.g. palmitic acid ethyl ester), whereas fatty acids were detected only in dried leaf oil (e.g. palmitic acid). In general, fatty acid esters have low boiling points and easily volatilized, then they might be lost during the long period of air dry and fermentation. However, at the same time, some fatty acid esters might be hydrolyzed and caused the formation of fatty acids due to endogenous or microbial enzymes¹⁸. Hydrolysis of palmitic acid ethyl ester to palmitic acid was the example. As same as fatty acid ester, most of hydrocarbon compounds can easily volatile, then decreasing in the amounts of amyl vinyl carbinol, amyl ethyl carbinol, amyl ethyl ketone was observed. However, some increasing in the amount of amyl vinyl ketone was detected. Functional groups were the only difference among these compounds, and the highest oxidative degree was the functional group of the amyl vinyl ketone. Therefore, the increasing of this compound was possibly due to the oxidation of its derivatives in the fresh leaves.

Apocarotenoids were the known degradative

products derived from carotenoids by enzymatic and non-enzymatic oxidation. Increasing in their amounts after drying or fermentation was normally observed in many plant materials such as *Morus alba*, *M. nigra*¹⁹, black tea and Oolong tea²⁰. The result of this study was also in the same manner. After drying and fermentation, increasing in their proportion was observed. The relative amounts of apocarotenoids in the fresh leaf oil (4.9% in DB-5 and 1.7% in Carbowax 20M) were much increased in the dried leaf oil (11.9% in DB-5 and 12.1% in Carbowax 20M) than fermented leaf oil (5.7% in DB-5 and 2.8% in Carbowax 20M). Three significantly increasing compounds were beta-ionone, hexahydrofarnesyl acetone and farnesyl acetone C. Degradation mechanisms of carotenoids to form these three compounds have already been reported^{19,21}.

Biological activities of some major compounds of essential oils of *P. serratifolia* have been reported and supported the traditional usage of anticancer and the treatment of liver diseases. Linalool have been reported for antimicrobial, anti-inflammatory, anticancer, and antioxidant properties²². (*E*)-caryophyllene, palmitic acid, phytol, apocarotenoids and amyl vinyl carbinol possessed anticancer, and antioxidant activities²³⁻²⁷. beta-Myrcene was antiproliferative compound²⁸, whereas *p*-vinylanisole and methyl salicylate were shown to have antioxidant activity²⁹⁻³⁰. However, after drying or fermentation, amounts of some compounds increased and some compounds decreased. Therefore, biological activities of *P. serratifolia* leaves after drying or fermentation should be varied in potency from the fresh leaves and should be studied in more detail.

4. CONCLUSIONS

After drying and fermentation, a significant change in the chemical composition of essential oil of the leaves of *P. serratifolia* was detected. Some compounds disappeared, some new compounds occurred, and most compounds changed in their relative amounts. Dehydration, hydrolysis, and oxidation were suggested as the transformation reactions of some compounds. This result indicated the importance of the post-harvesting process on the quality of this herb. Bioactivity of its fresh, dried and fermented leaves would be further studied.

5. ACKNOWLEDGEMENT

We would like to thank Faculty of Pharmacy, Silpakorn University, Thailand, for providing chemical, glassware, equipment, and supporting laboratory facilities used this study.

Conflict of interest

None to declare.

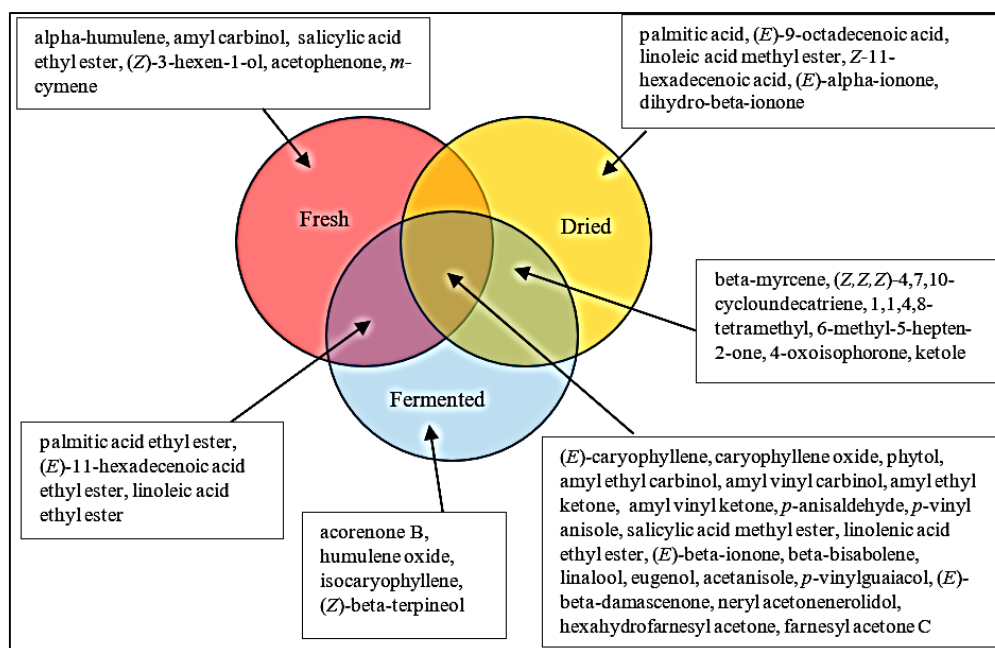


Figure 1. Comparison of some constituents of the essential oils of the fresh, dried and fermented *P. serratifolia* leaves.

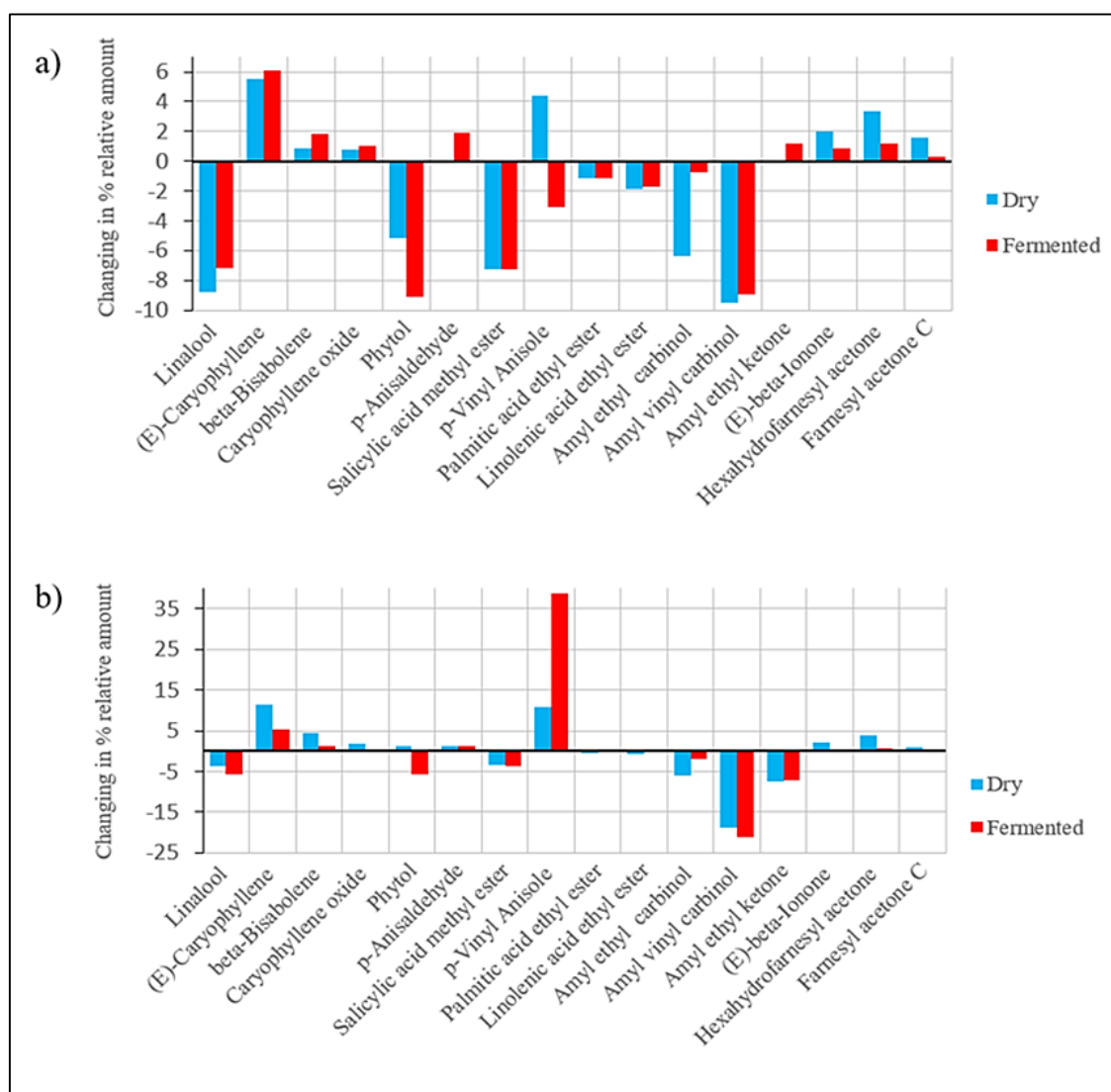


Figure 2. Changing in relative amounts of some constituents of the essential oils of the leaves of *P. serratifolia* after drying and fermentation, analysed by DB-5 (a) and Carbowax 20M (b) columns.

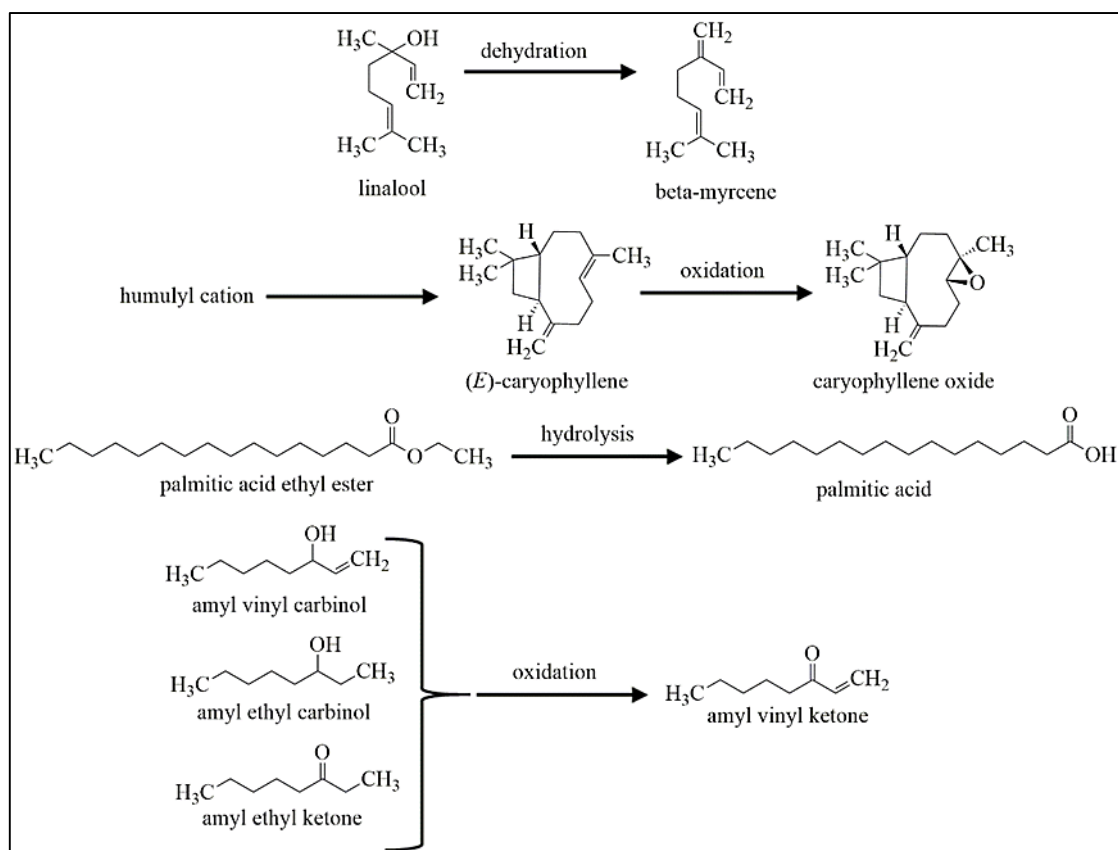


Figure 3. Proposed transformation mechanisms of some volatile compounds of the leaves of *P. serratifolia* after drying and fermentation.

Funding

None to declare.

Ethics approval

None to declare.

Article info:

Received April 17, 2020

Received in revised form July 28, 2020

Accepted December 9, 2020

REFERENCES

- Nurliana L, Musta R, Rudi L. Microencapsulation of essential oil from rogo plant (*Premna serratifolia* L.) as antibacterium *Escherichia coli*. Eng Sci Technol. 2018;7:314-23.
- de Kok R. The genus *Premna* L.(Lamiaceae) in the flora Malesiana area. Kew Bull. 2013;68(1):55-84.
- Atwolara-Odule OC, Oladosu IA. Comparison of chemical compositions of essential oils from the fresh and dried leaves of *Tapinanthus bangwensis* (Engl. and K. Krause) Danser [Loranthaceae]. Am J Essent Oil Nat Prod. 2016;4(3):31-3.
- Adeogun OO, Maroyi A, Afolayan AJ. Variation in the chemical composition of essential oils from *Artemisia afra* (Jacq) ex-wild leaf obtained by different methods and the effect of oil extracts on *Artemia salina* L. Trop J Pharm Res. 2018;17(3):519-28.
- Mirdha B, Naik S, Mahapatra S. Antimicrobial activities of essential oils obtained from fresh and dried leaves of *Ocimum sanctum* (L.) against enteric bacteria and yeast. Acta Hort. 2007;756:267-70.
- Hanaa AM, Sallam Y, El-Leithy A, Aly SE. Lemongrass (*Cymbopogon citratus*) essential oil as affected by drying methods. Ann Agric Sci. 2012;57(2):113-6.
- Baldermann S, Yang Z, Katsuno T, Tu VA, Mase N, Nakamura Y, et al. Discrimination of green, Oolong, and black teas by GC-MS analysis of characteristic volatile flavor compounds. Am J Anal Chem. 2014;5:620-32.
- Shou-liang C, Gilbert MG. Verbenaceae. In: Wu Z, Raven PH, editors. Flora of China, Volume 17: Verbenaceae through Solanaceae. Missouri Botanical Garden Press; 1994. p. 1-49.
- Munir AA. A taxonomic revision of the genus *Premna* L. (Verbenaceae) in Australia. J Adel Bot Gard. 1984;7(1):1-43.
- Leeratiwong C, Chantaranonthai P, Paton AJ. A synopsis of the genus *Premna* L.(Lamiaceae) in Thailand. Trop Nat Hist. 2009; 9(2):113-42.
- Adams RP. Identification of Essential Oil Components by Gas Chromatography/Mass Spectrometry. 4th ed. Allured Publishing Corporation: Carol Stream; 2007.
- National Institute of Standard and Technology Chemistry WebBook, SRD 69; 2018. [cited 2018 October]. Available from: <http://www.webbook.nist.gov>.
- Babushok V, Linstrom P, Zenkevich I. Retention indices for frequently reported compounds of plant essential oils. J Phys Chem Ref Data. 2011;40(4):1-15.
- Leffingwell JC, Alford E, Leffingwell D, Penn R, Mane S. Identification of the volatile constituents of Cyprian Latakia tobacco by dynamic and static headspace analyses. Leffingwell Rep. 2013;5(2):1-29.
- Nestl BM, Geinitz C, Popa S, Rizek S, Haselbeck RJ, Stephen R, et al. Structural and functional insights into asymmetric enzymatic dehydration of alkenols. Nat Chem Biol. 2017;13(3): 275-81.
- Yu F, Okamoto S, Nakasone K, Adachi K, Matsuda S, Harada H, et al. Molecular cloning and functional characterization of α -humulene synthase, a possible key enzyme of zerumbone biosynthesis in shampoo ginger (*Zingiber zerumbet* Smith). Planta. 2008;227(6):1291-9.
- Turek C, Stintzing FC. Stability of essential oils: a review. Compr Rev Food Sci F. 2013;12(1):40-53.

18. Mozafari AA, Vafae Y, Shahyad M. Phytochemical composition and *in vitro* antioxidant potential of *Cynodon dactylon* leaf and rhizome extracts as affected by drying methods and temperatures. *J Food Sci Tech*. 2018;55(6):2220-9.
19. Radulović NS, Miljković VM, Mladenović MZ, Nikolić GS. Essential oils of *Morus alba* and *M. nigra* leaves: Effect of drying on the chemical composition. *Nat Prod Commun*. 2017; 12(1):115-18.
20. Ho C-T, Zheng X, Li S. Tea aroma formation. *Food Sci Hum Wellness*. 2015;4(1):9-27.
21. Bonne T. Oxidation and thermal degradation of carotenoids. *J Oil Palm Res*. 1999;2:62-78.
22. Kamatou GP, Viljoen AM. Linalool-A review of a biologically active compound of commercial importance. *Nat Prod Commun*. 2008;3(7):1183-92.
23. Dahham SS, Tabana YM, Iqbal MA, Ahamed MB, Ezzat MO, Majid AS, et al. The anticancer, antioxidant and antimicrobial properties of the sesquiterpene β -caryophyllene from the essential oil of *Aquilaria crassna*. *Molecules*. 2015;20(7): 11808-29.
24. Ravi L, Krishnan K. Cytotoxic potential of *N*-hexadecanoic acid extracted from *Kigelia pinnata* leaves. *Asian J Cell Biol*. 2017;12:20-7.
25. Islam MT, Ali ES, Uddin SJ, Shaw S, Islam MA, Ahmed MI, et al. Phytol: A review of biomedical activities. *Food Chem Toxicol*. 2018;121:82-94.
26. Sharoni Y, Linnewiel-Hermoni K, Khanin M, Salman H, Veprík A, Danilenko M, et al. Carotenoids and apocarotenoids in cellular signaling related to cancer: A review. *Mol Nutr Food Res*. 2012;56(2):259-69.
27. Al-Fatimi M, Wurster M, Lindequist U. Chemical composition, antimicrobial and antioxidant activities of the volatile oil of *Ganoderma pfeifferi* Bres. *Medicines (Basel)*. 2016;3(2):10.
28. Blowman K, Magalhães M, Lemos MFL, Cabral C, Pires IM. Anticancer properties of essential oils and other natural products. *Evid Based Complementary Altern Med*. 2018(7);1-12.
29. Pereira DM, Valentão P, Pereira JA, Andrade PB. Phenolics: From chemistry to biology. *Molecules*. 2009;14:2202-11.
30. Oloyede GK. Toxicity, antimicrobial and antioxidant activities of methyl salicylate dominated essential oils of *Laportea aestuans* (Gaud). *Arab J Chem*. 2016;9:840-45.