

Review Article

Pharmacological treatment for the Novel Coronavirus Disease 2019 (COVID-19 Infection)

Nattapong Tidwong^{1,2*},
Preecha Montakantikul²,
Weerawat Manosuthi³

¹ Faculty of Pharmacy, Chiang Mai University,
Chiang Mai, Thailand

² Faculty of Pharmacy, Mahidol University,
Bangkok, Thailand

³ Bamrasnaradura Infectious Diseases Institute,
Nonthaburi, Thailand

*Corresponding author:
nattapong.tidwong@cmu.ac.th

KEYWORDS:

Novel coronavirus; COVID-19;
SARS-CoV-2; Antivirals

ABSTRACT

As of March 22, 2020, a total of 292,142 confirmed coronavirus disease 2019 (COVID-19) cases have been reported globally. Although there are currently no specific antiviral agents but all coronaviruses shared similar key elements of target for currently approved antiviral or new drug development. Several agents might be considered as a possible treatment based on the efficacy in SARS and MERS.

1. INTRODUCTION

In late December 2019, a series of pneumonia with unknown cause emerged in Wuhan, Hubei, China, with clinical presentation greatly resembling viral pneumonia. Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) was identified as the cause of this pneumonia which then was known as coronavirus disease 2019 (COVID-19)¹. As of March 22, 2020, a total of 292,142 confirmed cases have been reported globally. Identifying the treatment is crucial for the response to COVID-19 outbreak.

1.1. Characteristics of SARS-CoV-2

SARS-CoV-2 is an enveloped, positive-sense, single-stranded RNA betacoronavirus. The genomic characterization of SARS-CoV-2 suggested that this virus belongs to the subgenus Sarbecovirus, exhibited 79% and 50% identity to severe acute respiratory syndrome coronavirus (SARS-CoV) and middle east respiratory syndrome coronavirus (MERS-CoV), respectively². All coronaviruses shared similar key elements of target for currently approved antiviral or new drug development including two viral proteases (papain-like protease, 3C-like protease), non-structural protein (RNA-dependent RNA polymerase) and structural protein (such as spike glycoprotein)³. It is, therefore, reasonable to reconsider antiviral agents used in SARS and MERS for SARS-CoV-2. Here is a review of possible antiviral treatment.

1.2. Oseltamivir

Oseltamivir has long been used for treating influenza virus. It inhibits neuraminidase enzyme and prevents the release

of newly formed virions from the cell surface. Oseltamivir has also been used in coronavirus infection. During the emergence of MERS-CoV between 2003 and 2006 in Paris, oseltamivir in combination with effective antibiotics were considered as the empirical therapy for patients suspected with MERS⁴. Furthermore, recent report showed that almost all patients (89.9%) with confirmed SAR-CoV-2 infection in China received oseltamivir⁵. Considering *in vitro* activity, neuraminidase inhibitors, zanamivir and oseltamivir, has no *in vitro* antiviral activity against SARS-CoV even the concentration was as high as 1,000 and 10,000 μM , respectively⁶. It is likely because neuraminidase enzyme is not being used during coronavirus replication. However, anti-influenza virus might be beneficial since influenza virus infection was confirmed in about 30% in those patients⁴. Therefore, during the influenza season, oseltamivir might be considered as a part of medical treatment. Whether oseltamivir has a clinical benefit in SARS-CoV-2 should be confirmed by clinical trials (NCT04255017, NCT04261270).

1.3. Ribavirin

Ribavirin, a purine nucleoside analogue, inhibits enzymatic activity of Inosine-5'-monophosphate dehydrogenase (IMPDH), resulting in decrease intracellular guanosine triphosphate pools which then causes suppression in cellular DNA, mRNA and protein synthesis⁷. There were several studies showed antiviral activity of ribavirin against SARS-CoV and MERS-CoV. Ribavirin showed *in vitro* activity against two strains of SARS-CoV with the IC_{50} of $20 \pm 15 \mu\text{g/mL}$ (Frankfurt-1 strain) and $80 \pm 28 \mu\text{g/mL}$ (HKU39849 stain) but neither strain was completely inhibited even the concentration of ribavirin was as high as $100 \mu\text{g/mL}$ ⁸. Pharmacokinetic study in human after 1000 mg of ribavirin was administered intravenously resulting in mean plasma concentration of only $25 \mu\text{g/mL}$, suggested that plasma concentration after normal therapeutic dosing might not reach the inhibition level of SARS-CoV⁹. This finding was consistent with postmortem lung tissue sample collected from patient who died with a diagnosis of probable SARS, which still showed high viral loads (2.7×10^4 to 3.8×10^9 copies/g tissue) even the use of ribavirin or steroid¹⁰. This suggested that

ribavirin likely to have only a small beneficial effect in SARS. In addition, ribavirin has an activity against MERS-CoV *in vitro* with the IC_{50} quite high as $41.45 \mu\text{g/mL}$, however, the IC_{50} was reduced to $12 \mu\text{g/mL}$ when combined with interferon- $\alpha 2\text{b}$ (IFN- $\alpha 2\text{b}$) suggested that this combination may have benefit in MERS¹¹. Ribavirin combined with IFN- $\alpha 2\text{b}$ was then studied in the rhesus macaque model. Comparing to untreated macaque, treated macaque exhibited significantly better clinical outcomes including improved clinical parameters, better radiographic evidence of pneumonia, lower mean lung tissue viral load ($0.81 \log$ median tissue culture infectious dose (TCID_{50}) equivalents per gram tissue lower, $P=0.0428$), and reduced systemic and local pro-inflammatory markers¹². Unfortunately, the retrospective cohort study in critically ill patients with MERS suggested that the treatment with ribavirin in combination with recombinant interferon (rIFN- $\alpha 2\text{a}$, rIFN- $\alpha 2\text{b}$, or rIFN- $\beta 1\text{a}$) was not associated with 90-day mortality (adjusted odd ratio [aOR] 1.03, 95% confidence interval [95% CI] 0.73 to 1.44, $P=0.87$) or with more rapid MERS-CoV RNA clearance (aOR 0.65, 95% CI 0.30 to 1.44, $P=0.29$)¹³.

Recently, *in vitro* study showed that ribavirin also has an activity against SARS-CoV-2 with the EC_{50} of $109.5 \mu\text{M}$ ¹⁴ ($26.74 \mu\text{g/mL}$), which this concentration may achieved by normal therapeutic dosing. However, the efficacy in COVID-19 patients still need to be confirmed by clinical trials (NCT 04276688, NCT04306497).

1.4. Lopinavir/ritonavir

Lopinavir/ritonavir, a boosted protease inhibitor, was initially developed for a treatment of human immunodeficiency virus (HIV) infection but it can also inhibit the SARS-CoV, 3C-like protease (3CLpro) *in vitro* (IC_{50} $50 \mu\text{M}$)¹⁵ resulting in termination of viral replication. An open-labelled which compared the efficacy of treatment between a combination of lopinavir/ritonavir-ribavirin and ribavirin alone in SARS patients. An adverse outcome of acute respiratory distress syndrome (ARDS) or death within 21 days was significantly decreased in the combination group (absolute risk 26.4%, 95% CI 16.8 to 36.0, $P<0.001$). In addition, multiple logistic regression with adjusted baseline lactate dehydrogenase (LDH) also suggested that

treatment with lopinavir/ritonavir was independently associated with better outcome (aOR 0.076, 95% CI 0.01 to 0.589, $P=0.014$). Based on these studies, lopinavir/ritonavir is appeared to be a promising agent in SARS. In terms of MERS-CoV, lopinavir can inhibit viral replication *in vitro* with the EC_{50} of 8 μM ¹⁷, which is within the range of plasma concentration achievable by normal therapeutic dosing in HIV patients (8 to 24 μM)¹⁸. It also improve clinical, radiological and pathological outcomes in marmosets infected with MERS-CoV compared to untreated animals¹⁹. A clinical study on the efficacy of lopinavir/ritonavir in combination with IFN- β 1b in MERS is still on going (NCT02845843)²⁰.

In SARS-CoV-2, although there is lack of *in vitro* data, lopinavir/ritonavir (200/50 mg) 2 capsules twice a day can be considered as antiviral treatment base on certain benefits of lopinavir/ritonavir against SARS-CoV and MERS-CoV (weak recommendation)²¹. Recently, a randomized, controlled, open-label trial, which compared the efficacy of treatment between lopinavir/ritonavir plus standard care and standard care alone in patients with severe COVID-19 found that lopinavir/ritonavir treatment did not significantly accelerate clinical improvement (hazard ratio for clinical improvement, 1.24; 95% CI 0.90 to 1.72), reduce mortality (19.2% vs. 25.0%; difference, -5.8%; 95% CI, -17.3 to 5.7), or diminish throat viral RNA detectability compared to standard care alone²². Due to the several limitations of those study (eg. underpowered, not blinded), COVID-19 patients were further recruiting in clinical trials for efficacy evaluation of lopinavir/ritonavir and the results are still awaiting. (NCT04252885, NCT04261907, NCT04255017). In addition, other protease inhibitors, darunavir/cobicistat, was also evaluated for the treatment of COVID-19 in China (NCT04252274). The dosage regimen used in COVID-19 trials were lopinavir/ritonavir (200 mg/50 mg tablet) two tablets twice daily for 7-14 days or darunavir/cobicistat (800 mg/150 mg) 1 tablet once daily for 7 days.

1.5. Remdesevir (GS-5734)

Remdesevir, a nucleoside analog prodrug, can competitively incorporates with viral RNA-dependent RNA polymerase (RdRp), resulting in RNA synthesis inhibition²³. *In vitro* data showed

that remdesevir can effectively inhibit replication of both SARS-CoV (IC_{50} 0.069 μM) and MERS-CoV (IC_{50} 0.074 μM) and also improve lung viral load, clinical sign of lung disease, and respiratory function in mouse model²⁴. In addition, compared to lopinavir/ritonavir, remdesevir has superior anti-MERS-CoV activity, with lower EC_{50} *in vitro* (0.09 μM vs 8.5 μM), better improvement of pulmonary function, reduced lung viral load, and reduced severe lung pathology in transgenic mouse model²⁵. Data in MERS-CoV rhesus macaque model also provided additional benefits. Prophylaxis with remdesevir at 24 hours prior to inoculation completely inhibited MERS-CoV replication in lung tissue and effectively prevented clinical disease associated with MERS-CoV. Therapeutic treatment after 12 hours inoculation also reduced in clinical signs, reduced virus replication in the lungs, and decreased presence and severity of lung lesions²⁶. According to low *in vitro* inhibitory concentration (IC_{50} , EC_{50}) and vivid outcomes in animal model against both SARS-CoV and MERS-CoV, remdesivir was considered as a promising agent to be evaluated for the efficacy in SARS-CoV-2. Recent *in vitro* study showed that remdesevir efficiently inhibits SARS-CoV-2 with low EC_{50} of 0.77 μM ¹⁴ and one patient infected with SARS-CoV-2 was clinically improved after treatment with remdesevir²⁷. Several clinical trials are recruiting patients to evaluate the clinical efficacy of remdesivir in COVID-19 (NCT04252664, NCT04257656) using the dosage regimen of 200 mg loading dose on the first day, following by 100 mg intravenous once-daily maintenance dose for 9 days.

1.6. Sofosbuvir

Sofosbuvir, a nucleotide analog, was originally approved for the treatment of hepatitis C virus (HCV) infection. It inhibits HCV NS5B RNA-dependent RNA polymerase, which is essential for viral replication, and acts as a chain terminator²⁸. Recently, the molecular docking experiment using SARS-CoV-2 RNA dependent RNA polymerase (RdRp) model identified tight binding of sofosbuvir and ribavirin to the coronavirus RdRp²⁹, suggesting the possibility of using sofosbuvir and ribavirin in COVID-19 infection. However, the efficacy of this combination should be further confirmed by clinical trial.

1.7. Favipiravir (T-705)

Favipiravir, a purine nucleoside analog, was originally developed and approved for the treatment of influenza viruses in Japan. Unlike oseltamivir, favipiravir is phosphoribosylated to an active form, favipiravir ribofuranosyl-5B-triphosphate, which is then recognized as a purine nucleotide by RdRp, and inhibits its activity. Favipiravir inhibits *in vitro* replication of wide range of influenza viruses and many other RNA viruses including arenaviruses, bunyaviruses, flaviviruses, alphaviruses, paramyxoviruses, and norovirus family³⁰. Favipiravir also suppressed replication of Zaire ebolavirus (EBOV) *in vitro* with IC₅₀ of 67 µM and can induced rapid virus clearance, reduced biochemical parameters of disease severity, and prevented a lethal outcome in mice model³¹. Based on these data, favipiravir has been proposed to treat patients infected with EBOV. Considering SARS-CoV-2, recent study showed that favipiravir has *in vitro* antiviral activity with the EC₅₀ of 61.88 µM, which was higher than one with chloroquine and remdesivir¹⁴. Patients hospitalized with COVID-19 are being recruited for clinical trial evaluating the efficacy of favipiravir (ChiCTR2000029600, ChiCTR2000029544, NCT 04303299, NCT04310228, NCT04273763). The dosage regimen used in COVID-19 trials were mostly based on the dosing approved for influenza treatment, for example, 1,600 mg twice daily on the first day, following by 600 mg twice daily for 6 days.

1.8. Chloroquine

Chloroquine is an antimalarial agent with additional antiviral effect. It inhibits viral infection by increasing endosomal pH required for virus/cell fusion, as well as interfering with the glycosylation of cellular receptor of SARS-CoV. Chloroquine effectively inhibits SARS-CoV *in vitro* with the IC₅₀ of 8.8 µM³². It also has an *in vitro* activity against MERS-CoV (IC₅₀ of 4.1 µM)³³ and SARS-CoV-2 (EC₅₀ of 1.13 µM)¹⁴. Chloroquine has an additional immunomodulatory activity by suppressing the production and release of tumor necrosis factor (TNF) and interleukin 6 (IL-6), which mediate the inflammatory complications of several viral diseases. In addition, hydroxychloroquine, a less toxic derivative of chloroquine,

also has an *in vitro* activity against SARS-CoV-2 with the EC₅₀ of 4.06 µM, which was higher when compared to chloroquine (3.81 µM)³⁴. According to those *in vitro* data, chloroquine and hydroxychloroquine are being evaluated for the efficacy in COVID-19 patients (ChiCTR2000029609, ChiCTR2000029741, NCT04303299). The dosage regimen used in COVID-19 trials were chloroquine 500-1,000 mg per day for 7-10 days or hydroxychloroquine 400 mg per day for 5-10 days.

1.9. Umifenovir (Arbidol)

Umifenovir, also known as Arbidol, was approved in Russia and China for the prophylaxis and treatment of influenza A and B infection. It interacts with haemagglutinin (HA) to stabilize it against the low pH transition to its fusogenic state and consequently inhibit HA-mediated membrane fusion during influenza infection³⁵. For other viruses, it intercalates into membrane lipid leading to inhibition of membrane fusion between virus particle and plasma membrane, and between virus particle and the membrane of endosome³⁶. Umifenovir showed *in vitro* active against numerous DNA and RNA viruses including Lassa virus (IC₅₀ of 1.42 µM) and Ebola virus (IC₅₀ of 2.83 µM)³⁷. Interestingly, it also exhibited *in vitro* activity against SARS³⁸. Based on these data, clinical trials have been initiated to test umifenovir in patients infected with SARS-CoV-2 (NCT04260594, NCT04254874, NCT04255017, NCT04252885)

1.10. Interferon

Interferon (IFN) inhibit viral infection by directly interfering with viral replication by inducing both innate and adaptive immune response to infection³⁹. IFN-α and IFN-β have an *in vitro* activity against SARS-CoV and MERS-CoV^{6, 40}. Among IFN subtype, IFN-β showed the strongest *in vitro* inhibition against MERS-CoV with the lowest IC₅₀ of 1.3 U/mL compared to other subtype (IFN-α2a, IFN-α2b, IFN-γ, and IFN-universal)⁴¹. Various combinations of IFN-α or IFN-β with other antivirals such as ribavirin and/or lopinavir/ritonavir have been used to treat patients with MERS. Overall, combination treatments consisting of interferons and ribavirin did not consistently

improve outcomes in MERS¹³. However, the randomized double blinded trial on the efficacy of IFN- β 1b combination with lopinavir/ritonavir in MERS patients is still ongoing (NCT02845843). In addition, based on efficacy in SARS-CoV and MERS-CoV, combination of antiviral and various subtype of interferon (eg. PegIFN- α 2b, IFN- α 1b, IFN- β 1a) were studied for the efficacy in COVID-19 patients (ChiCTR2000029387, NCT04293887, NCT04254874).

2. CONCLUSIONS

Currently, there are no specific antiviral agents for COVID-19. Several agents might be considered as a possible treatment based on the efficacy in SARS and MERS. Due to the limited data, the most likely promising agent should be remdesivir (GS-5734) due to its low inhibitory concentration and satisfied therapeutic outcomes in SARS and MERS. However, favipiravir (T-705) and chloroquine might be considered. In addition, apart from non-structural protein, spike glycoprotein-targeted and host-targeted agents is currently in development, suggested other possible options to treat COVID-19. However, the efficacy and safety of all proposed anti-SARS-CoV-2 agents should be further confirmed by clinical studies.

Article info:

Received February 25, 2020

Received in revised form March 22, 2020

Accepted March 26, 2020

REFERENCES

1. Lu H, Stratton CW, Tang YW. Outbreak of pneumonia of unknown etiology in Wuhan, China: The mystery and the miracle. *J Med Virol*. 2020;92(4):401-2.
2. Lu R, Zhao X, Li J, Niu P, Yang B, Wu H, et al. Genomic characterisation and epidemiology of 2019 novel coronavirus: implications for virus origins and receptor binding. *Lancet*. 2020;395(10224):565-74.
3. Zumla A, Chan JFW, Azhar EI, Hui DSC, Yuen K-Y. Coronaviruses — drug discovery and therapeutic options. *Nat Rev Drug Discov*. 2016;15(5):327-47.
4. Bleibtreu A, Jaureguierry S, Houhou N, Boutolleau D, Guillot H, Vallois D, et al. Clinical management of respiratory syndrome in patients hospitalized for suspected Middle East respiratory syndrome coronavirus infection in the Paris area from 2013 to 2016. *BMC Infect Dis*. 2018;18(1):331.
5. Wang D, Hu B, Hu C, Zhu F, Liu X, Zhang J, et al. Clinical Characteristics of 138 Hospitalized Patients With 2019 Novel Coronavirus-Infected Pneumonia in Wuhan, China. *JAMA*. 2020;323(11):1061-9.
6. Tan ELC, Ooi EE, Lin C-Y, Tan HC, Ling AE, Lim B, et al. Inhibition of SARS coronavirus infection in vitro with clinically approved antiviral drugs. *Emerg Infect Dis*. 2004;10(4):581-6.
7. Cameron CE, Castro C. The mechanism of action of ribavirin: lethal mutagenesis of RNA virus genomes mediated by the viral RNA-dependent RNA polymerase. *Curr Opin Infect Dis*. 2001;14(6):757-64.
8. Saijo M, Morikawa S, Fukushi S, Mizutani T, Hasegawa H, Nagata N, et al. Inhibitory effect of mizoribine and ribavirin on the replication of severe acute respiratory syndrome (SARS)-associated coronavirus. *Antivir Res*. 2005;66(2-3):159-63.
9. Koren G, King S, Knowles S, Phillips E. Ribavirin in the treatment of SARS: A new trick for an old drug? *CMAJ : Can Med Assoc J*. 2003;168(10):1289-92.
10. Mazzulli T, Farcas GA, Poutanen SM, Willey BM, Low DE, Butany J, et al. Severe acute respiratory syndrome-associated coronavirus in lung tissue. *Emerg Infect Dis*. 2004;10(1):20-4.
11. Falzarano D, de Wit E, Martellaro C, Callison J, Munster VJ, Feldmann H. Inhibition of novel beta coronavirus replication by a combination of interferon-alpha2b and ribavirin. *Sci Rep*. 2013;3:1686.
12. Falzarano D, de Wit E, Rasmussen AL, Feldmann F, Okumura A, Scott DP, et al. Treatment with interferon-alpha2b and ribavirin improves outcome in MERS-CoV-infected rhesus macaques. *Nat Med*. 2013;19(10):1313-7.
13. Arabi YM, Shalhoub S, Mandourah Y, Al-Hameed F, Al-Omari A, Al Qasim E, et al. Ribavirin and Interferon Therapy for Critically

- Ill Patients With Middle East Respiratory Syndrome: A Multicenter Observational Study. *Clin Infect Dis.* 2019. ciz544
14. Wang M, Cao R, Zhang L, Yang X, Liu J, Xu M, et al. Remdesivir and chloroquine effectively inhibit the recently emerged novel coronavirus (2019-nCoV) in vitro. *Cell Res.* 2020;30(3): 269-71.
 15. Wu CY, Jan JT, Ma SH, Kuo CJ, Juan HF, Cheng YS, et al. Small molecules targeting severe acute respiratory syndrome human coronavirus. *Proc Natl Acad Sci U S A.* 2004; 101(27):10012-7.
 16. Chu CM, Cheng VCC, Hung IFN, Wong MML, Chan KH, Chan KS, et al. Role of lopinavir/ritonavir in the treatment of SARS: initial virological and clinical findings. *Thorax.* 2004; 59(3):252-6.
 17. de Wilde AH, Jochmans D, Posthuma CC, Zevenhoven-Dobbe JC, van Nieuwkoop S, Bestebroer TM, et al. Screening of an FDA-approved compound library identifies four small-molecule inhibitors of Middle East respiratory syndrome coronavirus replication in cell culture. *Antimicrob Agents Chemother.* 2014;58(8):4875-84.
 18. Lopez Aspiroz E, Santos Buelga D, Cabrera Figueroa S, Lopez Galera RM, Ribera Pascuet E, Dominguez-Gil Hurla A, et al. Population pharmacokinetics of lopinavir/ritonavir (Kaletra) in HIV-infected patients. *Ther Drug Monit.* 2011;33(5):573-82.
 19. Chan JF, Yao Y, Yeung ML, Deng W, Bao L, Jia L, et al. Treatment With Lopinavir/Ritonavir or Interferon-beta1b Improves Outcome of MERS-CoV Infection in a Nonhuman Primate Model of Common Marmoset. *J Infect Dis.* 2015;212(12):1904-13.
 20. Arabi YM, Allothman A, Balkhy HH, Al-Dawood A, AlJohani S, Al Harbi S, et al. Treatment of Middle East Respiratory Syndrome with a combination of lopinavir-ritonavir and interferon-beta1b (MIRACLE trial): study protocol for a randomized controlled trial. *Trials.* 2018;19(1):81.
 21. Jin Y-H, Cai L, Cheng Z-S, Cheng H, Deng T, Fan Y-P, et al. A rapid advice guideline for the diagnosis and treatment of 2019 novel coronavirus (2019-nCoV) infected pneumonia (standard version). *Mil Med Res.* 2020;7(1):4.
 22. Cao B, Wang Y, Wen D, Liu W, Wang J, Fan G, et al. A Trial of Lopinavir-Ritonavir in Adults Hospitalized with Severe Covid-19. *N Engl J Med.* 2020. [Epub ahead of print]
 23. Tchesnokov EP, Feng JY, Porter DP, Götte M. Mechanism of Inhibition of Ebola Virus RNA-Dependent RNA Polymerase by Remdesivir. *Viruses.* 2019;11(4):326.
 24. Sheahan TP, Sims AC, Graham RL, Menachery VD, Gralinski LE, Case JB, et al. Broad-spectrum antiviral GS-5734 inhibits both epidemic and zoonotic coronaviruses. *Sci Transl Med.* 2017;9(396): eaal3653.
 25. Sheahan TP, Sims AC, Leist SR, Schafer A, Won J, Brown AJ, et al. Comparative therapeutic efficacy of remdesivir and combination lopinavir, ritonavir, and interferon beta against MERS-CoV. *Nat Commun.* 2020;11(1):222.
 26. de Wit E, Feldmann F, Cronin J, Jordan R, Okumura A, Thomas T, et al. Prophylactic and therapeutic remdesivir (GS-5734) treatment in the rhesus macaque model of MERS-CoV infection. *Proc Nat Acad Sci USA.* 2020: 201922083.
 27. Holshue ML, DeBolt C, Lindquist S, Lofy KH, Wiesman J, Bruce H, et al. First Case of 2019 Novel Coronavirus in the United States. *N Engl J Med.* 2020;382(10):929-36.
 28. Gotte M, Feld JJ. Direct-acting antiviral agents for hepatitis C: structural and mechanistic insights. *Nat Rev Gastroenterol Hepatol.* 2016;13(6):338-51.
 29. Elfiky AA. Anti-HCV, nucleotide inhibitors, repurposing against COVID-19. *Life Sci.* 2020; 248:117477.
 30. Furuta Y, Gowen BB, Takahashi K, Shiraki K, Smeets DF, Barnard DL. Favipiravir (T-705), a novel viral RNA polymerase inhibitor. *Antivir Res.* 2013;100(2):446-54.
 31. Oestereich L, Ludtke A, Wurr S, Rieger T, Munoz-Fontela C, Gunther S. Successful treatment of advanced Ebola virus infection with T-705 (favipiravir) in a small animal model. *Antivir Res.* 2014;105:17-21.
 32. Vincent MJ, Bergeron E, Benjannet S, Erickson BR, Rollin PE, Ksiazek TG, et al. Chloroquine is a potent inhibitor of SARS coronavirus infection and spread. *Virol J.* 2005;2:69.
 33. Liang R, Wang L, Zhang N, Deng X, Su M, Su Y,

- et al. Development of Small-Molecule MERS-CoV Inhibitors. *Viruses*. 2018;10(12):721.
34. Liu J, Cao R, Xu M, Wang X, Zhang H, Hu H, et al. Hydroxychloroquine, a less toxic derivative of chloroquine, is effective in inhibiting SARS-CoV-2 infection in vitro. *Cell Discov*. 2020; 6(1):16.
35. Leneva IA, Russell RJ, Boriskin YS, Hay AJ. Characteristics of arbidol-resistant mutants of influenza virus: implications for the mechanism of anti-influenza action of arbidol. *Antivir Res*. 2009;81(2):132-40.
36. Villalain J. Membranotropic effects of arbidol, a broad anti-viral molecule, on phospholipid model membranes. *J Phys Chem B*. 2010;114(25):8544-54.
37. Hulseberg CE, Fénéant L, Szymańska-de Wijs KM, Kessler NP, Nelson EA, Shoemaker CJ, et al. Arbidol and Other Low-Molecular-Weight Drugs That Inhibit Lassa and Ebola Viruses. *J Virol*. 2019;93(8):e02185-18.
38. Khamitov RA, Loginova S, Shchukina VN, Borisevich SV, Maksimov VA, Shuster AM. [Antiviral activity of arbidol and its derivatives against the pathogen of severe acute respiratory syndrome in the cell cultures]. *Vopr Virusol*. 2008;53(4):9-13.
39. Zorzitto J, Galligan CL, Ueng JJ, Fish EN. Characterization of the antiviral effects of interferon-alpha against a SARS-like coronavirus infection in vitro. *Cell Res*. 2006;16(2): 220-9.
40. de Wilde AH, Raj VS, Oudshoorn D, Bestebroer TM, van Nieuwkoop S, Limpens RW, et al. MERS-coronavirus replication induces severe in vitro cytopathology and is strongly inhibited by cyclosporin A or interferon-alpha treatment. *J Gen Virol*. 2013;94(Pt 8):1749-60.
41. Hart BJ, Dyal J, Postnikova E, Zhou H, Kindrachuk J, Johnson RF, et al. Interferon-beta and mycophenolic acid are potent inhibitors of Middle East respiratory syndrome coronavirus in cell-based assays. *J Gen Virol*. 2014; 95(Pt 3):571-7.